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Application of Pulsed Electric Field to Process Milk

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Abstract: Milk is a nutritious food consumed by people of all ages. It is highly nutritious and thus is highly perishable unless and until preserved by some heat treatment like sterilization, ultra-heat treatment or boiling. In the present article, scope of utilizing pulsed electric field (PEF) has been discussed as an emerging technique of preservation to provide safe, fresh and shelf-stable milk with minimum or no nutrient losses. It covers details about the equipment, process and its effects on concerned microorganisms, enzymes and nutrients.

Keywords: Pulsed electric field, food, milk, sterilization, pasteurization.

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Introduction

Milk supplies energy and essential nutrients to nourish body. However, it is highly perishable and is, therefore, normally preserved with the application of heat. Various thermal techniques like pasteurization and sterilization are widely employed to process milk which however, also result in undesirable browning, development of cooked flavour, reduction of nutritional properties like losses in vitamins particularly B-complex and impairment of rennet-ability. To overcome such effects of thermal processing techniques on milk, several new and emerging methods of milk preservation are under scientific exploration. In the present paper pulsed electric field (PEF) is discussed to understand whether it can be used as a promising processing technique for milk. Indian dairy is growing at a CAGR of 6.4% and has produced 198.4 million metric tonnes of milk in 2019-20 maintaining its number one position in World Milk Production. Exploitation of novel technologies like PEF to process and extend the shelf-life of milk will offer new opportunities to the young Indians who are exploring dairy sector to venture into start-ups which in turn will give an added thrust to dairy industry.

In PEF, short pulses of high intensity electric fields (1-100 kV/cm) are applied for a period of microseconds to milliseconds to the food product flowing between two electrodes at ambient or moderately elevated temperatures. Depending on various process parameters, PEF treatment inactivates microorganisms and enzymes to varying degree. However, PEF brings minimum changes in the sensory attributes, nutrition of the treated food products and helps maintain 'fresh-like' quality (Upadhyay *et al.* 2019). PEF is considered as a superior method in comparison to thermal processing and preservation (Reineke *et al.* 2015). In India, Defence Food Research Laboratory (DFRL, Mysore) has conducted studies on the effect of PEF in combination with heat or sonication and have been able to extend the shelf-life of milk (flavoured) to 120 days at 5 °C and 90 days at ambient condition and that of curd to 180 Days at 5 °C and 21 days at ambient condition, respectively (Kumar *et al.* 2019).

In the early 1900s, ohmic heating (rise in temperature due to electrical resistance) was used to inactivate micro-organisms and to obtain pasteurized milk. Beattie and Lewis (1925) designed the equipment with 3000–4000 V to process milk electrically and Fetterman (1928) processed milk using the "Electro Pure Process", where milk at 70°C was passed through carbon electrodes in an electric heating chamber to inactivate *Mycobacterium tuberculosis* and *Escherichia coli*. In the last decade, research is focused on studying the potential of PEF to inactivate microorganisms and enzymes in milk due to the growing importance of dairy industry (Bendicho *et al.* 2002a; Sharma *et al.* 2014). Globally various research scientists are working on industrial applications of PEF in the different parts of the World.

2. PEF Equipment and Process

The main components of typical PEF equipment are a high voltage pulse generation system, a treatment chamber assembly (continuous or static), a pump for subjecting liquid food for example, milk, to enable continuous PEF treatment along with necessary monitoring and controlling devices and an aseptic packaging system to avoid post-processing contamination (Kumar *et al.* 2019). High voltage pulse generator supplies electrical energy of selected voltage. Heat exchangers can be installed for pre-heating and cooling of the food before and after PEF treatment. (Sharma *et al.* 2014). Processing time can be calculated by multiplying the number of pulses by effective pulse duration (Upadhyay *et al.* 2019). Barbosa-Canovas *et al.* (1999) reported several designs of PEF treatment chambers; out of which parallel plates, coaxial and co-linear configurations are the most commonly used. Bellebna *et al.* (2017) have tried

different configurations comprising of stainlesssteel electrodes and square shape treatment chamber made of Plexiglas to achieve savings in electricity as well as higher processing output.

3. Microbial Inactivation in Milk by PEF

Knowledge about the mechanism of microbial inactivation helps in designing efficient PEF equipment. Based on microbial and physicochemical studies using phospholipid vesicles and planar bilayers model systems, PEF induced microbial inactivation is believed to be due to dielectric breakdown and electroporation of the cell membrane (Ho and Mittal, 2000). In 1967, Sale and Hamilton described membrane damage as the direct cause of cell inactivation. Some authors have suggested that it starts by electroporation wherein there is electrical breakdown of the cell wall and cytoplasm contents leak out resulting in cell death. The electroporation theory suggests that the main effect of an electric field on microbial cells is to increase the membrane permeability due to membrane compression, cell poration and cell inactivation resulting from osmotic imbalance across the cell membrane (Tsong, 1990). Large pores can be obtained by increasing the intensity of the electric field and pulse duration or by reducing the ionic strength of the medium (Schoenbach *et al.* 1997).

The effectiveness of PEF treatment to inactivate microorganisms depends on various factors such as process parameters (electric field intensity, number of pulses, pulse shape, frequency, and duration of pulse), product parameters (composition, conductivity, pH, and water activity), and microbial characteristics (type of microorganism, growth conditions, growth phase, and recovery conditions). The research activities reported in the literature have focused on the impact of PEF treatment on microbial and enzymatic inactivation in milk or SMUF. The SMUF is a salt solution with composition similar to milk ultrafiltrate. It was proposed by Jeness and Koops, (1962) and is now widely used in dairy-related research (Bendicho et al. 2002). Various studies published on treatment of milk by PEF have proven this technology as an effective method for the inactivation of moulds, yeasts and vegetative bacterial cells. The microorganisms inactivated by PEF belong to the major Gram +ve and Gram -ve bacteria. Various researchers have reported 1 to 6 logs inactivation of different strains of *E. coli* (pathogenic and non-pathogenic) in milk (UHT, skim, whole, partially skim), egg pulp, pea soup, apple juice, SMUF, 0.1% NaCl saline, phosphate buffer (pH 7.0) and sodium alginate. Dutreux et al. (2000) studied effect of PEF treatment of E. coli and L. innocua suspended in pasteurised skim milk and in phosphate buffer (inlet and outlet temperatures: 17°C and 37°C; flow rate: 0.5 L/min; frequency: 3 Hz; field intensity: 41 kV/cm). They found leakage of cytoplasm due to the partially broken cell membrane. Level of microbial inactivation has been found to be mainly dependent on the electric field strength, number of pulses applied during the process and treatment time (Table 1). Rowan et al. (2001) investigated the viability of Mycobacterium paratuberculosis cells suspended in 0.1% (w/v) peptone water and in sterilised cow's milk as influenced by PEF. While PEF treatment at 5°C reduced the cells only by 1.6 logs, treatment at 50°C with 2,500 pulses of 5 Hz and 30 kV/cm reduced the number of viable *M. paratuberculosis* cells by approximately 5.3 logs in 0.1% peptone water and 5.9 logs in cow's milk. Evrendilek and Zhang, (2005) reported effects of pulse polarity and "pulse delaying time" (the time elapsed between two consecutive crests passing a given point) on the inactivation of *E. coli* O157: H7 in apple juice and skim milk (field strengths of 31 and 24 kV/cm, respectively; pulse delaying times: 3 to 1430 μ s). The pH and electrical conductivity for apple juice (3.7±0.24 and 2.3 mS/cm) and for skim milk $(6.7\pm0.65 \text{ and } 6.2\pm3.4 \text{ mS/cm})$, was found to be related to the significant difference observed in E. coli O157:H7 numbers in skim milk between mono (1.27 logs) and bipolar (1.96 logs) pulses, but not in apple juice (2.6 and 2.63 logs, respectively) at the pulse delaying time of 20 μ s. To achieve a higher level of microbial inactivation in milk, PEF can be used in combination with heat or organic acids. Molina *et al.* (2005) investigated the shelf life of various PEF-treated skim milks at room temperature or conventional heating at 60 or 65°C for 21 s as well as the combination of PEF treatment and heat or organic acids (acetic or propionic acids) on total number of aerobic bacteria (including *Pseudomonas fluorescens*). In all the studies, PEF treatment with exponential decaying pulses, field intensities of 30 to 50 kV/cm, pulse frequency of 4 Hz and treatment temperature of 40 to 65°C in combination with organic acids had a greater effect on inactivation of microorganisms than PEF alone or combined with mild temperature. In an attempt to further extend the shelf-life of HTST pasteurised milks by PEF treatment Sepulveda et al. (2005) subjected pasteurised milk to PEF treatment immediately after pasteurisation and after 8 days storage at 4°C using field intensity of 35 kV/ cm and 2 pulses of 2.3 µs duration each. The final temperature was 65°C with a residence time of less than 10 s. It was shown that the application of PEF immediately after pasteurisation could extend the shelf life of milk up to 60 days at 4°C, while PEF processing after 8 days storage resulted in a longer shelf life of 78 days due to further eradication of enteric and psychrotrophic bacteria by PEF.

S. No.	Type of Milk	PEF treatment	Microorganism studied	Log reduction	Reference
1.	Fruit juice– whole or skim milk bever- ages	35 kV/cm, for 1800 μs	Listeria innocua	5 Log cycle	Salvia Trujillo et al. (2011)
2.	Milk	40 kV/cm	Listeria innocua	4.3 Log cycle	Guerrero-Bel- trán at el. (2010)
3.	UHT Milk	31 kV/cm	Pseudomonas	>5 Log cycle	Craven <i>et al.</i> (2008)
4.	UHT Full Fat Milk	30 – 60 kV/ cmfor 26 – 210 μs	E. Coli	8 Log cycle	Shin <i>et al.</i> (2007)
			Pseudomonas fluorescens	8 Log cycle	
			Bacillus stearothermo- philus	3 Log cycle	
5.	Raw Skim Milk, UHT Skim Milk	35 kV/cm, 188 μs	Pseudomonas fluorescens, Lactococcus lactis, Bacil- luscereus	0.3 – 3 Log cycle	Michalac <i>et al.</i> (2003)
6.	Cow Milk	30 kV/cm, 2500 pulses	Mycobacterium tubercu- losis	5.9Log cycle	Rowan <i>et al.</i> (2003)

Table 1: Effect of PEF on Milk Microflora

To explore the potential of PEF technology. some studies have been undertaken to understand the effect of PEF in combination with presence of food grade antimicrobials like nisin and lysozyme (Mittal and Griffiths, 2005). Pol *et al.*, 2000 reported a synergistic effect on subjecting vegetative cells of *B. cereus* to combination of nisin and PEF treatment with 1.8 logs extra reduction in B. cereus numbers than the sum of the reductions obtained from the individual treatments. Calderon *et al.* (1999) combined PEF treatment with nisin addition to inactivate *Listeria innocua* in skim milk. The selected field intensities (and temperatures) were 30 (22°C), 40 (28°C) and 50 (34°C) kV/cm and the number of pulses applied were 10.6, 21.3 and 32, respectively. *Listeria innocua* count was reduced to 2, 2.7 and 3.4 logs after exposure to the field intensities of 30, 40 and 50 kV/cm, respectively in presence of 10 IU nisin/mL while at 100 IU nisin/mL under the same PEF treatment conditions the reduction increased to 2.5, 3 and 3.8 logs. The increase in microbial reduction was attributed to the additive effect of nisin on PEF treatment.

3.1. Effects of PEF on Milk enzymes

The impact of PEF on inactivation of enzymesis not so clearas some cases report a high level of inactivation while in other cases no effect was observed (Loey *et* al. 2002; Martin-Belloso et al. 2005; Yu et al. 2012). The effects of PEF treatment on activities of various milk enzymes including alkaline phosphatase, lipases, lactoperoxidase and proteases commonly present in milk or SMUF has been reported by several researchers (Castro et al. 2001; Riener et al. 2009; Sharma et al. 2014). Sharma et al. (2014) reported a reduced activity of enzymes plasmin (12%) and xanthine oxidase (32%) on PEF treatment of 26.1 kV/cm for 34 μ s in combination with preheating at 55 °C for 24 s. Riener *et al.* (2009) found a reduction in activity of lipase, alkaline phosphatase and protease of fresh bovine milk by 14%, 29% and 37%, respectively on the application of 35 kV/cm for 75 μ s. Enzyme inactivation requires a more severe PEF treatment than that needed for inactivating microorganisms (Ho *et al.* 1997). The higher the electric field intensity and temperature, the greater reduction in enzyme activity is achievable. Barsoti and Cheftel, (1999) related unfolding, denaturation, breakdown of covalent bonds and oxidation-reduction reactions in the protein structure to enzyme inactivity.

3.2. Effects of PEF on the Functionality of Milk Nutrients

Thermal method of preserving milk affects the physico-chemical and functional properties of milk proteins and fat globules. The processing parameters adopted during PEF treatment determines to large extent the changes that will bring denaturation in protein or modify the milk fat structure or nutrient content, for example, rise in temperature. The content of protein and fat greatly affect the yield and physical characteristics of the products made from the milk therefore, it is important to understand the effect of PEF technology on protein and fat as it can have a direct impact on product economics and consumer liking. Upadhyay et al. (2019) reported that PEF treatment of milk at low intensities does not significantly affect the proteins, fats, vitamins and other milk nutrients. Thermal or non-thermal treatment of cheese milk has been known to affect the final properties of cheese directly or indirectly. Floury et al. (2006) reported that the PEF treatments (field intensities: 45 or 55 kV/cm; pulse widths: 500 and 250 square monopolar pulses, respectively), decreased the coagulation time. At a total treatment time of just 2.1-3.5 μ s, there was a significant drop in casein micelle size along with a decrease in milk viscosity and the coagulation properties were enhanced. Wüst *et al.* (2004) found that increasing the field strength decreased the strength of the cottage

cheese gel made from PEF-treated skim milk (bipolar square pulses of 2 μ s; field intensities: 25 and 28 kV/cm; pulse frequency: 200 and 400 Hz; and flow rate of 120 mL/min at a treatment temperature of less than 45°C). The yield of cottage cheese was marginally increased compared to cheeses made from raw or pasteurised skim milk. At a frequency of 400 Hz, the "raw milk" odour was also got removed. Yu *et al.* (2012) found better rennetability of PEF treated milk (treatment at 30kV/cm or less) in comparison to thermally treated milk (heat treated at 50 °C or less).

Unlike thermal processing, PEF treatment (30.76 to 53.84 kV/cm) of skim and whole milk at different temperatures of 20 °C, 30 °C, and 40 °C, brought just minor variations in pH, electrical conductivity, density, colour, solids-notfat (Bermudez-Aguirre *et al.*, 2011) and Reiner *et al.* (2009) reported not much effect of PEF treatment on retinol, vitamin E, thiamine or riboflavin content of raw milk. Hemar *et al.* (2011) found no significant change in the viscosity of concentrated milk (total solids 18%) on PEF treatment at 45 kV/cm for 20 μ s.

4. Limitations of PEF

Processing the milk products having air bubbles with PEF technology results in non-uniform treatment which could affect the operational safety. Further, milk products with suspended particulates could cause system malfunction. There should not be any clumps of particles and the maximum particle size in the milk must be smaller than the gap between the electrodes in the chamber for the uniform processing operation. Also, innovations and engineering advancements are required to adopt the PEF technology in its application to handle milk and milk products at large scale with maximum output and efficiency.

5. Conclusion

PEF, as a non-thermal process, does not largely alter the original composition of milk. It has the potential to serve as an alternative to traditional heat treatment of milk with the advantages of maintaining sensory and nutritional quality as of fresh produce. Intensive studies are required to assure the safety of PEF technology in achieving the desired level of enzymatic and microbial inactivation when used alone or in combination with other processing hurdles like heat, preservatives or acids. Further, in order to make PEF technology applicable in the dairy industry, large scale PEF systems will be required with the equivalent efficiency of currently available bench-top or pilot scale systems.

References

- Aronsson K, Lindgren M, Johansson BR and Ulf R. 2001. Inactivation of microorganisms using pulsed electric fields: the influence of process parameters on *E. coli*, *L. innocua*, *L. mesenteroides* and *S. cerevisiae*.Innovative Food Science and Emerging Technology. 2, 41-54.
- Barbosa-Caanovas GV, Gongora-Nieto MM, Pothakamury UR and Swanson BG. 1999. Design of PEF processing equipment. In: Preservation of foods with pulsed electric fields. San Diego, CA, USA: Academic Press. 20-46.
- Barbosa-Canovas GV, Pierson MD, Zhang QH and Schaffner DW 2000. Pulsed Electric Fields. Special Supplement: Kinetics of Microbial Inactivation for Alternative Food Processing Technologies. Journal of Food Science. 65, 65-68.
- Barsotti L and Cheftel JC 1999. Food processing by pulsed electric fields. II. Biological aspects. Food Reviews International. 15, 181-213.
- Beattie JM, and Lewis FC. 1925. The electric current (apart from the heat generated). A bacteriological agent in the sterilization of milk and other fluids. J. Hygiene, 24, 123–137.
- Bellebna Y, Bermmaki H, Semmak A, Chaker A and Tilmatine A. 2017. Study and analysis of new pulsed electric field treatment chamber configurations for food extraction. Turkish Journal of Electrical Engineering & Computer Sciences. 25: 4149 4159.
- Bendicho S, Barbosa-Canovas GV and Martin O. 2002. Milk processing by high intensity pulsed electric fields. Trends in Food Science & Technology, 13, 195-204.
- Bendicho S, Estela C, Fernandez-Molina JJ, Barbosa-Canovas GV and Martin O. 2002a. Effects of high intensity pulsed electric fields and thermal treatments on a lipase from Pseudomonas fluorescens, Journal of Dairy Science, Vol. 85, pp. 19-27.
- Bermudez-Aguirre D, Fernandez S, Esquivel H, Dunne PC and Barbosa-Canovas GV. 2011. Milk processed by pulsed electric fields: Evaluation of microbial quality, physicochemical characteristics, and selected nutrients at different storage conditions. J Food Sci. 76(5): 289-299.
- Calderon-Miranda ML, Barbosa-Canovas GV and Swanson BG. 1999. Transmission electron microscopy of Listeria innocua treated by pulsed electric fields and nisin in skimmed milk. International Journal of Food Microbiology. 51, 31-38.
- Castro A, Barbosa-Canovas GV and Swanson BG. 1993. Microbial inactivation of foods by pulsed electric fields. Journal of Food Processing and Preservation. 17, 47-73.
- Castro AJ, Swanson BG, Barbosa-Canovas GV and Zhang QH. 2001. Pulsed Electric Field Modification of Milk Alkaline Phosphatase. In G. V. Barbosa-Canovas, Q. H. Zhang, Pulsed Electric Fields in Food Processing-fundamental aspects and applications. Food preservation technology series Lancaster (PA): Technomic Publishing Co. pp. 65-82.
- Craven HM, Swiergon P, Ng S, Midgely J, Versteeg C, Coventry MJ and Wan J. 2008. Evaluation of pulsed electric field and minimal heat treatments for inactivation of

pseudomonads and enhancement of milk shelf-life. Innov Food Sci Emerg Technol. 9(2): 211-216.

- Dutreux N, Notermans S, Wijtzes T, Gongora-Nieto MM, Barbosa-Canovas GV and Swanson BG. 2000. Pulsed electric fields inactivation of attached and free-living Escherichia coli and Listeria innocua under several conditions, International Journal of Food Microbiology. 54, 91-98.
- Evrendilek GA and Zhang QH. 2005. Effects of pulse polarity and pulse delaying time on pulsed electric fields-induced pasteurisation of E. coli O157:H7. Journal of Food Engineering. 68, 271-276.
- Fetterman JC. 1928. The electrical conductivity method of processing milk. Agricultural Engineering. 4, 407–408.
- Floury J, Grosset N, Leconte N, Pasco M, Madec M and Jeantet R. 2006. Continuous raw skim milk processing by pulsed electric field at non-lethal temperature: effect on microbial inactivation and functional properties. Lait, 86, 43-57.
- Getchell BE' 1935. Electric pasteurization of milk. Agricultural Engineering, 16, 408–410.
- Guerrero-Beltrán JA, Sepulveda DR, Góngora-Nieto MM, Swanson B, and Barbosa-Cánovas GV. 2010. Milk thermization by Pulsed electric fields (PEF) and electrically induced heat. J Food Eng. 100(1): 56-60.
- Hemar Y, Augustin MA, Cheng LJ, Sanguansri P, Swiergon P and Wan J. 2011. The effect of pulsed electric field processing on particle size and viscosity of milk and milk concentrates. Milchwissenschaft. 66(2) 126.
- Ho SY and Mittal GS. 2000. High voltage pulsed electrical field for liquid food pasteurization. Food Reviews International, 16, 395-434.
- Ho SY, Mittal GS and Cross JD. 1997. Effects of high field electric pulses on the activity of selected enzymes, Journal of Food Engineering. 31, 69-84.
- Jeness R and Koops J. 1962. Preparation and properties of a salt solution which simulates milk ultrafiltrate, Netherlands Milk and Dairy Journal. 16, 153-164.
- Kinosita KJ and Tsong TY. 1977. Voltage induced pore formation and haemolysis erythrocytes, Biochimica et biophysica acta. 471, 227-242.
- Kumar R, Vijayalakshmi S, Kathiravan T and Nadanasabapati S. 2019. PEF processing of fruits, vegetables and their products. In: Non-thermal processing of foods. (Ed.) O. P. Chauhan, Francis and Taylor, CRC Press, London, UK. 107-127. ISBN 9781315233017
- Loey VA, Verachtert B and Hendrickx M. 2002. Effects of high electric fields pulses on enzymes. Trends in Food Science and Technology. 12, 94-102.
- Martin-Belloso O, Bendicho S, Elez-Martinez P and Barbosa-Canovas GV. 2005. Does high intensity pulsed electric fields induce changes in enzymatic activity, protein conformation, and vitamin and flavour stability In Novel Food Processing Technologies. (Eds. G.V. Barbosa-Canovas, M.S. Tapia, M.P. Cano) CRC Press, pp. 87-103. ISBN 9780429225055

- Michalac S, Alvarez VT, JLJi T and Zhang QH. 2003. Inactivation of selected microorganisms and properties of pulsed electric field processed milk. J Food Process Preserv. 27(2); 137-151.
- Mittal GS and Griffiths MW. 2005. Pulsed Electric Field Processing of Liquid Foods and Beverages. In: Emerging Technologies for Food Processing. (Ed.) Da-Wen Sun, London: Elsevier Academic Press, pp. 99-139. ISBN 978-0-12-676757-5
- Mohamed MEA and Eissa AHA. 2012. Pulsed Electric Fields for Food Processing Technology, Structure and Function of Food Engineering, Ayman Amer Eissa, Intech Open, ISBN 978-953-51-0695-1 DOI:10.5772/48678. Available from: https://www. intechopen.com/books/structure-and-function-of-food-engineering/pulsed-electricfields-for-food-processing-technology.
- Molina FJJ, Altunakar B, Bermusez-Aguirre D, Swanson BG, Barbosa-Canovas GV. 2005. Inactivation of *Pseudomonas fluorescens* in skim milk by combination of pulsed electric fields and organic acids. Journal of Food Protection, 68, 1232-1235.
- Moses BD. 1938. Electric pasteurization of milk. Agricultural Engineering, 12, 525–526.
- Pol IE, Mastwijk HC, Bartels PV and Smid EJ. 2000. Pulsed-electric field treatment enhances the bactericidal action of nisin against Bacillus cereus. Applied and Environmental Microbiology. 66, 428-430.
- Pothakamury UR, Barbosa-Canovas GV, Swanson, BG and Spence KD. 1997. Ultrastructural changes in Staphylococcus aureus treated with pulsed electric fields, Food Science and Technology International. 3, 113-121.
- Reineke K, Schottroff F, Meneses N, *et al.* 2015. Sterilization of liquid foods by pulsed electric fields–an innovative ultra–high temperature process. Front Microbiol. 6: 400.
- Reiner J, Noci F, Cronin DA, Morgan DJ and Lyng JG.2009. Effect of high intensity pulsed electric fields on enzymes and vitamins in bovine raw milk. Int J Dairy Technol 62(1) 1-6.
- Rowan JN, MacGregor SJ, Anderson JG, Cameron D and Farish O. 2001. Inactivation of Mycobacterium paratuberculosis by pulsed electric fields, Applied and Environmental Microbiology, 6, 2833-2836.
- Sale AJH and Hamilton WA. 1967. Effects of high electric fields on microorganisms I. Killing of bacteria and yeast. Biochimica et biophysica acta. 163, 34-43.
- Salvia-Trujillo L, Morales-de la Pena M, Rojas-Grau MA, and Martin-Belloso O. 2011. Microbial and enzymatic stability of fruit juice-milk beverages treated by high intensity pulsed electric fields or heatduring refrigerated storage. Food Control 22(10): 1639-1646.
- Schoenbach KH, Peterkin FE, Alden RW and Beebe SJ. 1997. The effect of pulsed electric fields on biological cells: Experiments and applications, IEEE Transactions on Plasma Science. 25, 284-292.

- Sepulveda DR, Gongora-Nieto MM, Guerrero JA and Barbosa-Canovas GV. 2005. Production of extendedshelf-life milk by processing pasteurised milk with pulsed electric fields.Journal of Food Engineering. 67, 81-86.
- Shamsi K and Sherkat F. 2009. Application of pulsed electric field in non-thermal processing of milk. Asian Journal of Food and Agro-Industry. 2 (3) 216-244.
- Sharma P, Oey I, Everett DW. 2014. Effect of pulsed electric field processing on the functional properties of bovine milk. Trends in Food Science & Technology. 35(2):87-101.
- Shin JK, Jung KJ, Pyun YR and Chung MS. 2007. Application of pulsed electric fields with square wave pulse to milk inoculated with E. coli, P. fluorescens, and B. stearothermophilus. Food Sci Biorec16(6): 1082-1084.
- Tsong TY. 1990. Reviews on electroporation of cell membranes and some related phenomena, Bioelectrochemistry and Bioenergetics. 24, 271.
- Upadhyay N, Kumar MCT, Sharma H, Borad S and Singh AK. 2019. Pulse electric field processing of milk and milk products. In: Non-thermal processing of foods. OP Chauhan(ed.), CRC Press, UK. 129 -143.ISBN 978-1-1380-3584-3.
- Vega-Mercado H, Martin-Belloso O, Chang FJ, Barbosa-Cánovas GV and Swanson BG. 1996. Inactivation of *Escherichia coli* and *Bacillus subtilis* suspended in pea soup using pulsed electric fields. Journal of Food Processing and Preservation, Vol. 20, pp. 501-510.
- Wüst R, Pearce R, Ortega-Rivas E and Sherkat F. 2004. Pulsed electric fields treatment of milk affects the properties of the cottage cheese gel. 9th International Congress on Engineering and Food, Abst. No 896: 87. 7-11 March, Montpellier, France.
- Yu LJ, Ngadi M and Raghavan V. 2012. Proteolysis of cheese slurry made from pulsed electric field treated milk. Food Bioprocess Technol., 5(1) 47-54.
- Zhang QH, Barbosa-Canovas GV and Swanson BG. 1995. Engineering aspects of pulsed electric fields pasteurisation, Journal of Food Engineering. 25, 175-256.